

Biocompatible Inorganic Fullerene-Like Molybdenum Disulfide Nanoparticles Produced by Pulsed Laser Ablation in Water

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There has been a lot of interest in the study of nanostructures due to their unique properties and potential applications in new nanotechnologies.^{1–5} Molybdenum disulfide (MoS₂), tungsten disulfide (WS₂), and layered inorganic compounds were found to have a comparable structure to carbon nanoparticles, *i.e.*, hollow cage structure, like carbon “onions”, and nanotubes and could form inorganic fullerene-like (termed IF) structures under appropriate conditions.^{6–11} MoS₂ and WS₂ nanostructures have distinctive properties (like excellent lubrication properties)¹² and wide ranges of promising applications, such as heterogeneous catalysis, hydrogen storage, and lithium–magnesium ion batteries, *etc.*^{9–11} The first potential medical application—alleviating friction in orthodontic wires—was recently demonstrated.^{12–14} Furthermore, self-lubricating friction-reduction coatings with possible biomedical potential based on MoS₂ and WS₂ nanoparticles with inorganic fullerene-like (IF) structure, such as coatings for artificial joints, have been suggested by Samorodnitsky-Naveh *et al.*¹⁴ They showed that stable and well-adhered cobalt + IF-WS₂ coating of NiTi substrates was obtained. Friction tests presented up to 66% reduction of the friction coefficient. Since NiTi alloy is widely used for many medical appliances, this unique friction-reducing coating could be implemented to provide better manipulation. In addition, biofunctionalization of WS₂ nanostructures has also been achieved by Tahir *et al.*¹⁵ They have used a polymeric ligand containing the tetradentate nitrilotriacetic acid (NTA) groups with dual functionality to anchor functional groups to the surface sulfur layer and the

ABSTRACT We report on the synthesis of inorganic fullerene-like molybdenum disulfide (MoS₂) nanoparticles by pulsed laser ablation (PLA) in water. The final products were characterized by scanning electron microscopy, X-ray diffraction, transmission electron microscopy, and resonance Raman spectroscopy, *etc.* Cell viability studies show that the as-prepared MoS₂ nanoparticles have good solubility and biocompatibility, which may show a great potential in various biomedical applications. It is shown that the technique of PLA in water also provides a green and convenient method to synthesize novel nanomaterials, especially for biocompatible nanomaterials.

KEYWORDS: colloids · inorganic fullerene-like · biomedical applications · nanoparticles · pulsed laser ablation · biocompatibility

linker groups for biofunctionalization through the immobilization of His-tagged protein (silicatein) to the functionalized chalcogenide surface. The protein retains its catalytic activity when immobilized on the surface of WS₂ nanotubes. Biofunctionalization of WS₂ nanostructures opens new opportunities for integrating this group of nanomaterials in composites, especially for immobilization of various proteins and biomolecules. All these progresses make metal chalcogenide nanoparticles good candidates for various biomedical applications, such as drug delivery (through controlled release of small molecules or proteins),¹⁵ cancer therapy, and medical-relevant coatings.

Before any material can be used for biomedical applications, its biocompatibility and toxicology must be evaluated. Preliminary toxicology tests of WS₂ nanoparticles revealed that the material showed no apparent toxic reaction after an oral administration test in rats and no sensitization in lymph nodes following a topical dermal application.^{16,17} More recent inhalation tests have shown that the material has no toxic effect in rats.¹⁸ Nevertheless more toxicity

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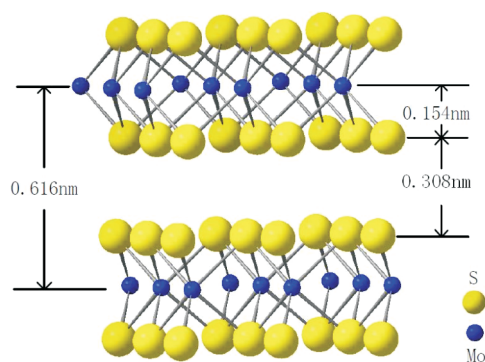


Figure 1. Schematic diagram of the 2H-MoS₂ structure. (The lattice parameters in figure were taken from ref 29).

and sensitivity tests need to be done in view of the emerging applications of these nanomaterials.

Up to now, much effort has been devoted to the synthesis of these novel nanostructures. Various methods have been reported,^{19–26} such as the gas-phase reaction between oxides and H₂S/H₂,^{20,21} decomposition of ammonium thiomolybdate or metal trisulfides,^{22,23} heating of MoS₂ powders,²⁴ and the catalyzed transport method, *etc.*²⁵ In this work, we use pulsed laser ablation (PLA) to prepare MoS₂ nanoparticles in deionized water. PLA in water is a relatively new method^{27,28} to prepare the metal and semiconductor nanoparticles, and it is also a potentially important method to produce nanoparticles which could be used in drug delivery and other biomedical applications. Compared with chemical methods, PLA in water can produce nanoparticles with several advantages: (1) Preparation is relatively simple and convenient; (2) the nanoparticle system is pure without other remnants from the chemical reactions; and (3) the products have high water solubility.

Here, we report on the synthesis of IF-MoS₂ nanoparticles by PLA of pressed pellets of 2H-MoS₂ powder in water. The final products were characterized by field emission scanning electron microscopy (FE-SEM), X-ray diffraction (XRD), transmission electron microscopy (TEM), and resonance Raman Spectroscopy, *etc.* Cell viability studies were conducted with the IF-MoS₂ nanoparticles and different human cells. The IF-MoS₂ nanoparticles were found to be nontoxic to these cells up to the stated concentrations.

RESULTS AND DISCUSSION

Figure 1 shows the structure of bulk MoS₂ crystal,²⁹ which contains a Mo layer sandwiched between two S layers with the Mo in a trigonal bipyramidal coordination. Strong covalent forces bind Mo and S atoms within a lamella, whereas adjacent lamellae interact through relatively weak van der Waals forces. The unit cell has hexagonal symmetry and includes two adjacent lamellae (termed 2H arrangement). This graphite-like structure is considered to be responsible for the lubricative

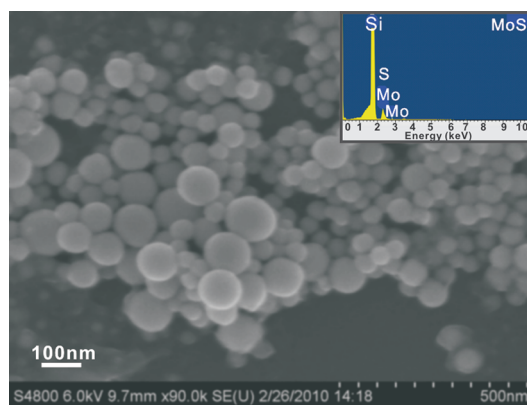


Figure 2. The FE-SEM image of MoS₂ nanoparticles and the EDS spectrum in the inset.

properties of these materials, because the weak interlamellar forces facilitate the shear when the direction of sliding is parallel to the planes of the material.

Figure 2 shows a SEM image of MoS₂ nanoparticles prepared by PLA in water. The starting material was a pressed pellet of MoS₂ powder. A series of analytical experiments was carried out in order to determine the composition and structure of the generated nanoparticles. One can see that the product consists of spherical nanoparticles. The average diameter of the MoS₂ nanoparticles is about 120 nm, as determined by dynamic light scattering (DLS) measurement (Supporting Information, Figure S2). Energy-dispersive X-ray spectrum (EDS, Horiba, installed on SEM) shows that the nanoparticles primarily contain Mo and S. (Figure 2 inset, the Si peak in the figure comes from the Si substrate).

XRD measurements were carried out to determine the structure and composition of the MoS₂ nanoparticles, and the result is shown in Figure 3. It can be observed that all the diffraction peaks in spectrum (a) can be indexed to MoS₂ with hexagonal structure (ICDD-JCPDS card no. 39-1492). In comparison, Figure 3b shows the XRD pattern of the 2H-MoS₂ powder. All the peaks in spectrum (b) can be assigned to the hexagonal 2H-MoS₂ with lattice constants $a = 3.160$, $c = 12.295$ Å (ICDD-JCPDS card no. 39-1492). One can see that the main peaks, *i.e.*, (002) and (006) in Figure 3a exhibit a small shift to lower angles compared to their corresponding ones in Figure 3b, which indicated about 0.3% lattice expansion. This shift can be attributed to the introduction of strain owing to curvature of the layers.³⁰

Figure 4 shows a typical high-resolution TEM (HRTEM) image of MoS₂ nanoparticles prepared by PLA in water. The layered structure of the walls forming these fullerene-like nanoparticles is quite clear from the image. The interlayer distance is about 6.18 Å, which corresponds to the d -spacing of MoS₂ (002) planes. The inset in Figure 4 is a line profile of the boxed region.

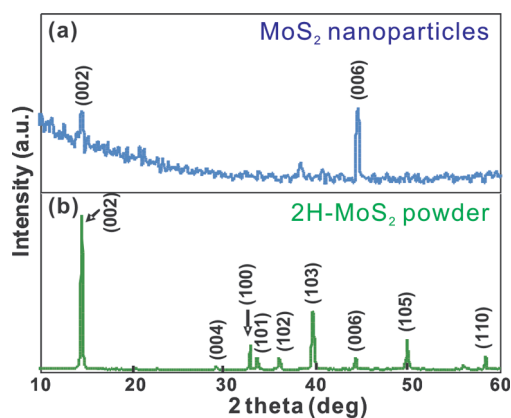


Figure 3. XRD patterns for: (a) MoS₂ nanoparticles and (b) 2H-MoS₂ powder.

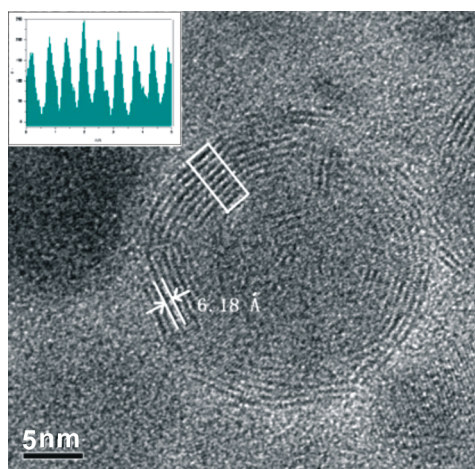


Figure 4. HRTEM image of MoS₂ nanoparticles.

Resonance Raman (RR) spectra of IF-MoS₂ nanoparticles and 2H-MoS₂ powder are shown in Figure 5, which were taken at room temperature using the 632.8 nm He–Ne laser excitation line. The modes shown in Figure 5 are listed in Table 1. In the RR spectrum of the 2H-MoS₂ powder, many peaks are observed. All the peaks are in good agreement with the reported MoS₂ single-crystal resonance Raman studies.^{31,32} In the RR spectrum of IF-MoS₂ nanoparticles, the most intense peak is centered at 405 cm⁻¹. This mode is assigned to the A_{1g}(Γ) mode.^{31,32} In the higher frequency region of the RR spectrum, peaks are observed at 593 and 638 cm⁻¹, which were assigned to the second-order processes: E_{2g}¹(M) + LA(M) and A_{1g}(M) + LA(M), respectively. In the lower frequency region of the RR spectrum, there are two peaks at 175 and 378 cm⁻¹, which were assigned to the A_{1g}(M) – LA(M) and E_{2g}¹(Γ), respectively. All the RR peaks of MoS₂ nanoparticles in Figure 5 shift to lower frequencies compared to their bulk ones, which is a typical phenomenon observed in MoS₂ nanostructures.^{32–34} (The peak at 520 cm⁻¹ comes from the Si substrate.)

One can also see from Figure 5 that the most intense peak in the RR spectrum of the 2H-MoS₂ powder is

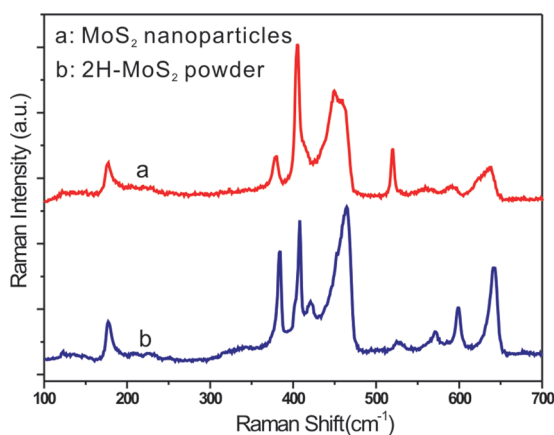


Figure 5. RR spectra of MoS₂ nanoparticles and 2H-MoS₂ powder.

TABLE 1. Raman Peaks Observed in MoS₂ Samples at Room Temperature and Corresponding Symmetry Assignments^a

MoS ₂ powder	IF-MoS ₂	symmetry assignment
176	175	A _{1g} (M) – LA(M)
384	378	E _{2g} ¹ (Γ)
408	405	A _{1g} (Γ)
421		
	450	
464	463	2 × LA(M)
524		E _{1g} (M) + LA(M)
571		2 × E _{1g} (Γ)
602	593	E _{2g} ¹ (M) + LA(M)
643	638	A _{1g} (M) + LA(M)

^a All peak positions are in cm⁻¹. LA refers to longitudinal acoustic, Γ and M refer to Γ and M points in the electronic-band structure diagrams.

centered at 464 cm⁻¹. This mode has been previously assigned by Stacy *et al.*³¹ to the second-order 2 × LA(M) mode. In the spectrum of MoS₂ nanoparticles, this peak is actually split into two peaks at 450 and 463 cm⁻¹, suggesting that this feature is a superposition of two peaks. Figure S3a, Supporting Information, shows that the fitted data and Lorentzian functions are used to deconvolute the feature into two peaks. The relative intensity, $P(\text{low peak})/P(\text{high peak})$, is larger for the sample of nanoparticles than that for the sample of the 2H-MoS₂ powder, which is in agreement with the results of Frey *et al.*³²

In light of these results, the formation mechanism of fullerene-like MoS₂ can be discussed. This process involves the interaction between pulsed laser radiation and the target source material. Stresses produced by laser-induced thermal shock were not relieved and could eventually lead to cracking and flake detachment, *i.e.*, “exfoliation”.³⁵ Laser exfoliation caused the delamination of nanosize layer structured MoS₂ materials. The ejected MoS₂ thin flakes and fragments were hit by shock waves produced during subsequent pulses, causing further diminution of particles. SEM images of the

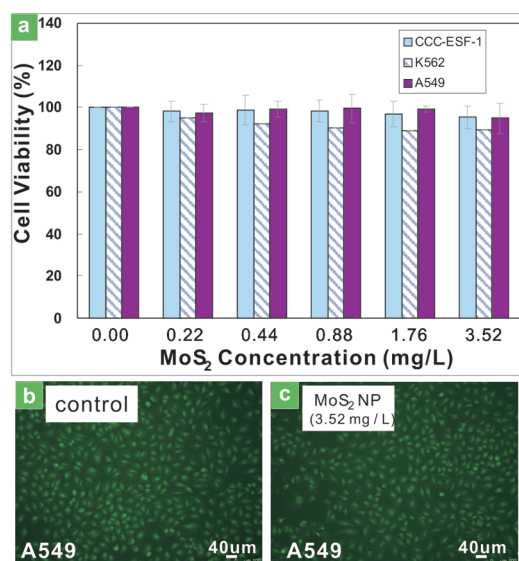


Figure 6. (a) Cell viability of different cells (A549, K562, and CCC-ESF-1) after incubating in increasing concentrations of MoS₂ nanoparticles, as determined by the MTT test. (b and c) fluorescent microscopy images of untreated control A549 cells and the cells treated with MoS₂ nanoparticles (3.52 mg/L) for 48 h, respectively.

starting MoS₂ powder and the IF-MoS₂ nanoparticles produced by PLA in water for 10 min are shown in Figure S5, Supporting Information. One can notice from that the sizes of those MoS₂ nanoparticles are larger than those ablated for 20 minutes, which also supports this explanation.

The use of nanoparticles *in vitro* and *in vivo* requires that they are biocompatible. The cytotoxic behavior of MoS₂ nanoparticles on cells was examined by the MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) assay. CCC-ESF-1 cell (human embryonic epidermal fibroblast cells) viability was assessed 48 h after exposure to different concentrations of MoS₂ nanoparticles. Control experiments were conducted in a similar manner without the presence of the MoS₂ nanoparticles. As shown in Figure 6a, cell viability data indicated that the MoS₂ nanoparticles did not significantly affect CCC-ESF-1 cell proliferation up to 3.52 mg/L particle concentration. In order to compare different cell viability in the presence of MoS₂ nanoparticles, we have tested two other different human cells: A549 cells

(lung adenocarcinoma cells) and K562 cells (leukemic cells). The results showed that the MoS₂ nanoparticles were reasonably nontoxic and biocompatible up to the given concentrations.

To study further the effect of MoS₂ nanoparticles on cell viability, we labeled A549 cells with acridine orange (Sigma Aldrich) and studied the cell morphologies under a fluorescent microscope. A549 cells were cultured in 96-well plates and incubated with MoS₂ nanoparticles with various concentrations, as discussed above. Figure 6b shows the fluorescent microscopy images of untreated control A549 cells and the cells treated with MoS₂ nanoparticles (3.52 mg/L) for 48 h. In acridine orange-stained cells, the nucleolus fluorescence was bright green, but the cytoplasm was a little dim. There was no obvious change of the morphologies after treatment with the MoS₂ nanoparticles for 2 days even at the highest concentration. There was nevertheless a little decrease in the cell numbers, which was similar to the case for the untreated cells. In conclusion we found that the MoS₂ nanoparticles were nontoxic to the cells at the given concentrations. It is interesting to compare cytotoxic effects of MoS₂ nanoparticles with those of other nanomaterials, such as single-walled carbon nanotube (SWCNT) particles. It was found by Manna *et al.*³⁶ that cell survival started to decrease after exposure to low concentration (0.15 mg/L) of SWCNT particles. Thus, MoS₂ nanoparticles seem to have a very good biocompatibility with the above human cells.

CONCLUSION

In summary, we prepared fullerene-like MoS₂ nanoparticles using PLA in water. The structure of the products was characterized by SEM, EDS, XRD, TEM, and Raman spectroscopy techniques. A mechanism for the nanoparticle synthesis was proposed. Cell viability studies show that the as-prepared MoS₂ nanoparticles have good solubility and biocompatibility, which may have an important impact on variety of applications, including various biomedical applications, such as medical friction-reduction coatings, drug delivery, and cancer therapy. The technique of PLA in water also provides a green and convenient method to synthesize novel crystalline materials, especially for biocompatible nanomaterials.

MATERIALS AND METHODS

Synthesis of MoS₂ Nanoparticles. Synthesis of MoS₂ nanoparticles was carried out by using PLA in water. The MoS₂ powder (Sigma Aldrich, 99.99% pure) was pressed into a target pellet (diameter of 15 mm) using a press system for infrared spectroscopy (a pressure of 12 MPa, 2 min). The pellet was placed at the bottom of a quartz cell with 5 mL redistilled deionized water. A Nd:YAG pulsed laser (532 nm, 5 Hz, pulse duration between 6 and 9 ns and pulse energy of 40–50 mJ), was focused on the MoS₂ pellet for ablation. The ablation was typically done for duration of 20 min. During the course of the ablation,

the colloidal solution was continuously stirred by a magnetic stirrer.

Characterization. The MoS₂ nanoparticles were filtered with 0.45 μm disposable filters and then characterized by FE-SEM (Hitachi S-4800), EDS (Horiba), TEM (Tecnai G220 ST), RR spectroscopy (Renishaw, the excitation wavelength was 632.8 nm), DLS (Malvern Zetasizer Nano ZS), and XRD (Bruker D8). The concentrations of the MoS₂ nanoparticle colloid solution used for the cell viability studies were measured with optical emission spectroscopy of inductively coupled plasma ICP-OES (Perkin-Elmer optima 5300DV).

Cell Culture. A549 and K562 cells were cultured in RPMI 1640 medium (Thermo Scientific) (CCC-ESF-1 cells were cultured in DMEM/high glucose medium) with 10% (V/V) fetal bovine serum (FBS) and 100 IU/mL penicillin and 100 μ g/mL streptomycin, respectively. The culture plates were incubated at 37 °C in a humidified incubator containing 5% CO₂.

MTT Assay for Cell Viability Test. The cell viability was tested using MTT assay, which is based on the mitochondrial conversion of tetrazolium salt. The A549 and K562 cells were seeded in 96-well microplates (Costar, Corning, NY) at a density of 8000 and 1.0×10^4 cells/well, respectively, in 100 μ L RPMI 1640 medium containing 10% FBS. CCC-ESF-1 cells were seeded at a density of 1.0×10^4 cells/well in DMEM/high-glucose medium. After attachment for 24 h, the cells were incubated with MoS₂ nanoparticles at various concentrations (0, 0.22, 0.44, 0.88, 1.76, and 3.52 mg/L) for another 48 h in a final volume containing 150 μ L medium, and then the medium was removed (except for experiments of K562 cells). After that, 150 μ L of fresh medium and 10 μ L of MTT (5 mg/mL in PBS) were added to each well, and the culture plates were incubated at 37 °C and 5% CO₂ for 4 h. After removal of the medium, 150 μ L of dimethyl sulfoxide (DMSO) was added to each well to dissolve the dye. The absorbance at 492 nm was measured using a microplate reader. Each data point was derived from three parallel samples.

Cell Viability Assay by Fluorescent Microscopy. The A549 cells were cultured in 96-well plates and mixed with MoS₂ nanoparticles, as discussed above. After treatment, the cells were labeled by incubation with acridine orange (Sigma Aldrich) at a concentration of 100 μ g/mL for 20 min at 37 °C in the dark. After incubation, the cells were washed twice with ice-cold complete medium and examined under a fluorescent microscope (Leica, Germany). Three sections were selected and examined randomly for each sample. Excitation was performed by an argon ion laser operating at 488 nm, and the emitted fluorescence was collected through a 515 nm pass filter.

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Supporting Information Available: Schematic diagram of the experimental setup, DLS of MoS₂ nanoparticles, a line-shape analysis of the 440–470 cm⁻¹ feature in the resonance Raman spectra of MoS₂ nanoparticles and powders, a HRTEM image of the MoS₂ nanoparticles, and FE-SEM images of MoS₂ powder and MoS₂ nanoparticles produced by PLA in water for 10 min. This material is available free of charge via the Internet at <http://pubs.acs.org>.

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